

Review Article

Cyperus spp.: A Review on Phytochemical Composition, Biological Activity, and Health-Promoting Effects

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Cyperaceae are a plant family of grass-like monocots, comprising 5600 species with a cosmopolitan distribution in temperate and tropical regions. Phytochemically, *Cyperus* is one of the most promising health supplementing genera of the Cyperaceae family, housing ≈950 species, with *Cyperus rotundus* L. being the most reported species in pharmacological studies. The traditional uses of *Cyperus* spp. have been reported against various diseases, *viz.*, gastrointestinal and respiratory affections, blood disorders, menstrual irregularities, and inflammatory diseases. *Cyperus* spp. are known to contain a plethora of bioactive compounds such as α -cyperone, α -corymbolol, α -pinene, caryophyllene oxide, cyperotundone, germacrene D, mustakone, and zierone, which impart pharmacological properties to its extract. Therefore, *Cyperus* sp. extracts were preclinically studied and reported to possess antioxidant, anti-inflammatory, antimicrobial, anticancer, neuroprotective, antidepressive, antiarthritic, antiobesity, vasodilator, spasmolytic, bronchodilator, and estrogenic biofunctionalities. Nonetheless, conclusive evidence is still sparse regarding its clinical applications on human diseases. Further studies focused on toxicity data and risk assessment are needed to elucidate its safe and effective application. Moreover, detailed structure-activity studies also need time to explore the candidature of *Cyperus*-derived phytochemicals as upcoming drugs in pharmaceuticals.

1. Introduction

Immemorially, human societies have been using herbs and their products as sources of medicine, nutrition, and industrial applications [1]. As an example of the role of plant species in human life, in ancient Egypt, the first paper was made from papyrus (*Cyperus papyrus* L.), a species of the Cyperaceae family [2]. Cyperaceae includes grass-like monocots, comprising around 5600 species and 100 genera, and the family is widespread on all continents with the exception of Antarctica. The second largest genus in this family is *Cyperus*, with ~950 species [3]. *Cyperus* spp. are most commonly known as weeds, despite some cultures using them for medicinal purposes and as a source of food [4]. *Cyperus* spp. predominantly exist in the wetlands throughout the globe in the tropical regions and act as source of primary productivity. The tubers, shoots, and fruits of this species are produced in larger quantities and act as a source of food for amphibians and aquatic animals [5].

The traditional use of *Cyperus* plants has been reported from all over the world as a remedy against various human ailments [6] including treatment of stomach and bowel disorders, as diuretic, digestant, and lactodepurant purposes. The plant extracts also act as a selective drug for the treatment of bronchitis, blood disorders, menstrual irregularities, amenorrhea, diarrhea, dysentery, and inflammatory diseases [7]. Interestingly and despite *Cyperus* including more than 950 species, the three most commonly reported species are purple nutsedge (*Cyperus rotundus* L.), yellow nutsedge (*Cyperus esculentus* L.), and *C. papyrus*. *Cyperus rotundus* is the most well-known species of *Cyperus* in South Asia, a perennial weed that grows best in high-moisture soil and reproduces easily through rhizomes and tubers [7]. This species is indigenous to the tropical and subtropical parts of the Old World, and despite the fact that it can be found detrimental in cultivated fields, it has several beneficial uses as medicine since ancient times [8]. *Cyperus rotundus* rhizomes and tubers are mentioned in Oriental traditional medicine to treat fever, digestive disorders, and menstrual irregularities in several countries including China, India, Iran, and Japan [9, 10].

Cyperus esculentus L. is an edible perennial grass-like plant native to the Old World. This species exists widely throughout tropics and subtropics of North America [11]. The earliest records of its use dated back to predynastic times about 6000 y ago in North America and Egypt; however, its different varieties are mostly found in Southern Europe, South-Middle East, and Africa [12]. It has been also considered as a foodstuff since ancient times, especially in ancient Egypt. It is a crop of early domestication and was regarded important with the other crops of the Nile Valley. Its dry tubers have been found in tombs from predynastic times about 6000 y ago. *Cyperus esculentus* tubers were roasted and used as a sweetmeat in Egypt during the ancient times [12]. *Cyperus esculentus* is widely cultivated for its edible tubers, called earth almonds or tigernuts [13], which are consumed as a popular snack in Africa and for making a sweet milk-like beverage, horchata de chufa, commonly consumed in Spain and other European and Latin-American countries

[14]. Tigernut is a rich source of protein and minerals making the beverage highly nutritious (phosphorus and potassium) [12].

Cyperus papyrus L. is an aquatic sedge mostly known for its use in the preparation of the paper by the traditional Egypt, Greek, and Roman civilizations. Paper made from dried, pressed, and woven strips of culm pith had been used since 3500 BC by ancient civilizations in the Egypt and the Mediterranean Basin. It was the only widespread recording medium until the 8th century in Europe [15]. Other species of the *Cyperus* family include *Cyperus compressus* L., *Cyperus javanicus* Houtt., and *Cyperus monocephalus* Roxb. (*Cyperus cephalotes*). For instance, *C. compressus* is a grass-like plant and is widely distributed across the tropical and subtropical regions of the world. In India, the powdered roots of *C. compressus* have long been used in traditional medicine by the Santhal tribes to treat intestinal helminthic infections [16]. Examples of folk medicinal and edible uses of *Cyperus* spp. reported from different parts of the world are briefly shown in Table 1.

Taken together, the multiple potentialities reported so far for the most widely exploited *Cyperus* spp. were considered. This review is the first of its kind that gives a comprehensive discussion on the recent findings related to chemical composition, biological activities, and pharmacological effects of such promissory naturally occurring matrices. The safety and toxicity effects of the *Cyperus* sp. extracts are also considered in the scope of the manuscript. The diagram showing various components discussed in the review are presented in Figure 1.

2. Chemical Composition

The Cyperaceae family is one of the largest flowering plant families and is ranked the third largest monocot family after Orchidaceae and Poaceae [46]. A rising number of studies have highlighted that the multiple potentialities of the species of this family as medicines are attributed to the presence of several bioactive constituents. For example, the cypriol, isolated from *Cyperus scariosus* R.Br. essential oil, is present in various perfumes and medicines. In fact, cypriol's ambery, balsamic, spicy, warm, and woody features make it highly demanded in perfume industry [47]. In addition, the essential oil is also present in various other species of *Cyperus*, such as *C. articulatus* L., *C. rotundus*, and *Cyperus maculatus* Boeckeler [48]. Summarization of phytochemicals present in the major six species of the *Cyperus* genus is summarized in Table 2, and in the next subsections, a brief description of the most abundant phytochemicals in the recently investigated *Cyperus* spp. is also presented.

2.1. *Cyperus articulatus* L. *Cyperus articulatus* is a perennial herb with underground perennial rhizomes having scales which grade into culm leaves. They have exceptionally high photosynthesizing function compared to other plants and are also regarded as herbal switch plants as they are a reservoir of potentially useful drugs for the treatment metabolic disorders [49]. Various specific compounds isolated from *C.*

TABLE 1: Examples of folk medicinal uses of a selection of *Cyperus* species.

Plant species	Country/region	Plant part (s)	Traditional use	Instruction	Reference
<i>Cyperus rotundus</i> L.	North-West Himalaya/India	Roots	Skin diseases	Decoction prepared by burning and adding the ash of fresh leaves of <i>A. baccifera</i> (10 g) and <i>C. rotundus</i> roots (10 g) and fresh ginger (5 g) in sesame oil.	[7]
	Pakistan/India	Tubers	Diabetes	10–12 g of dry tuber powder administered daily twice for 2–3 months.	[17]
	India	Whole plant	Menstruation problem	Juice of the <i>Citrus maxima</i> fruit (100 ml) and 30 g dried powder of <i>C. rotundus</i> is taken once daily for a week.	[18]
	Tamil Nadu/India	Tubers	Snake bite	Paste of leaf and root bark of <i>Albizia amara</i> , root bark of <i>Jasminum angustifolium</i> , and tubers of <i>C. rotundus</i> is heated with oil and applied externally on affected places for 10 days.	[19]
	India	Roots/tubers	Urinary trouble-stone removal	Decoction of the plant is used.	[20, 21]
	India	Whole plant	Epilepsy	Plant decoction (10 ml) with 5 ml of honey is orally administered to treat epilepsy.	[22]
	India	Roots	Cholera	Roots are boiled with equal quantity of mint and given for cholera.	[23]
	India	Roots	Pimples	Roots along with turmeric and curd are made into a paste which is applied on the face for pimples and beautification	[23]
	India	Roots	Increase lactation	Paste of the roots is applied on breasts to increase lactation.	[23]
	North-West Himalaya	Roots	Intermittent fevers	The decoction prepared from 10 g of <i>C. rotundus</i> roots and 5 g of fresh ginger is used.	[24] Das, & Misra [25]
India	India	Tubers	Dermatitis	The decoction prepared from tuberous roots of <i>C. rotundus</i> and leaf of <i>Trichosanthes anguina</i> is taken orally to cure dermatitis.	[25]
	India	Tubers	Dysentery	The tuberous root of <i>C. rotundus</i> with other plants which are orally used to treat dysentery is taken in three doses to cure dysentery.	[25]
	India	Tubers	Indigestion disorders, stomachache	A powder was prepared from 10 g of tuber of <i>C. rotundus</i> , 10 g stem bark of <i>Holarrhena antidyserterica</i> , and 10 g of <i>Zingiber officinalis</i> after being sun dried. 30 g powder is given internally along with 250 ml of buttermilk twice daily till cure.	[26]
	India	Tubers	Vaginal discharge	Tubers crushed with <i>Abutilon indicum</i> leaves and sufficient quantity of <i>Cuminum cyminum</i> seeds; extract administered daily twice for three days.	[27] Jahan et al. [28]
	India	Whole plant	Loss of libido in men	Leaves of <i>Psidium guajava</i> , leaves of <i>Punica granatum</i> , and whole plants of <i>C. rotundus</i> are mixed, warmed, and macerated to obtain juice. 1/2 cup of the juice is taken with 10–15 drops of honey twice daily for 3 days.	[28]
India	India	Tubers	Constipation	1/2 cup of juice obtained from macerated tubers is taken three times daily.	[28]
	India	Whole plant	Bone fracture	Whole plant of <i>C. rotundus</i> and 7 slices of ginger are crushed and made into a paste. The paste is warmed and applied to fractures.	[29]

TABLE 1: Continued.

Plant species	Country/region	Plant part (s)	Traditional use	Instruction	Reference
	India	Tubers	Jaundice	Fresh rhizome with tuberous root of <i>C. rotundus</i> and fruits of <i>Phyllanthus emblica</i> are taken in equal quantities and ground. 2 spoonfuls of paste mixed in a glass of water are administered daily once for 8 days.	[30]
	India	Bark	Malaria	The decoction is prepared from a mixture of 200 g of rhizome of <i>Costus speciosus</i> , 200 g bark of <i>C. rotundus</i> , and 200 g bark of <i>Azadirachta indica</i> . 2–4 spoons of decoction were prescribed after meal for 15 days.	[31]
	India	Tubers	Bronchitis	Tubers of <i>C. rotundus</i> and leaves of <i>Tinospora cordifolia</i> with fruits of <i>Pergularia daemia</i> are ground. 2 spoons of paste with honey are orally administered twice daily for 30 days.	[32]
	China	—	Coughs	—	[33]
	Rarotonga	Tubers	Sore throat	Twenty to thirty tubers of <i>C. rotundus</i> and a handful of <i>Pandanus tectorius</i> bark which is crushed into the water of four green coconuts. Half the mixture is drunk hot, and the remainder cold. The treatment lasts for three days.	[33]
<i>Cyperus javanicus</i> Houtt.	Rarotonga	Leaves	Fractures/sprains	Leaves without flowers are pounded and squeezed into a small basin of water. The treatment lasts for three days.	[33]
	Rarotonga	Leaves	Irregular menstrual	Leaves with those of several other herbs.	[33]
<i>Cyperus brevifolius</i> (Rottb.) Hassk.	Malaysia	Tubers	Sore legs	—	[33]
<i>Cyperus kyllingia</i> Endl.	Rarotonga	Tubers	Oral thrush	Tubers of <i>C. kyllingia</i> , 4 <i>Aleurites moluccana</i> inside nuts, and a handful of the aerial roots of <i>Ficus prolixa</i> are pounded then squeezed through a cloth into a liter of water.	[33]
<i>Cyperus monocephalus</i> Roxb.	Philippines Tami Islands	Tubers Tubers	Dermatosis Ringworm	Decoction is prepared from tuberous root. Decoction of tubers prepared by adding lime.	[33] [33]
<i>Cyperus compressus</i> L.	India	Roots	Helminthiasis	Powdered roots orally administered.	[16]
<i>Cyperus articulatus</i> L.	Central Africa Republic	Tubers	Headache, migraine	Decoction is prepared from tuberous root.	[34]
<i>Cyperus pedunculatus</i> (R.Br.) J.Kern	West Africa	Stem and leaves	Diarrhea, kidney disease, fever, pain, and inflammations	Extract is made from the whole plant	[35]
<i>Cyperus nitidus</i> Lam.	South Africa	Rhizomes	Respiratory and digestive disorders	Extract is made from the rhizomes	[36]
<i>Cyperus sexangularis</i> Nees	South Africa	—	Asthma, fatigue, fever, pneumonia, and TB	—	[37]
<i>Cyperus sexangularis</i> Nees	South Africa	Roots	Antimicrobial, emollient, diuretic, stimulant, anthelmintic, and analgesic treatment	Extract is made from the roots	[38]
	East Africa	Roots	Various animal diseases	Extract is made from the roots	[39]

TABLE 1: Continued.

Plant species	Country/region	Plant part (s)	Traditional use	Instruction	Reference
<i>Cyperus kilimandscharicus</i> Kük.					
<i>Cyperus latifolius</i> Poir.	East Africa	Roots	Tuberculosis and related ailments	Extract is made from the roots	[40]
<i>Cyperus maculatus</i> Boeck.	West Africa	Tubers	Cattle worms	—	[41]
<i>Cyperus natalensis</i> Hochst.	South Africa	Roots	Treatment of gynaecology and obstetric complaints	Decoction is prepared from the roots	[42]
<i>Cyperus erectus</i> (Schumach.) Mattf. & Kük.	South Africa	—	Reduces foot swelling	Ground plant is used for the medicinal purposes	[43]
<i>Cyperus mundii</i> (Nees) Kunth	Madagascar	—	Treatment of evacuation of the placenta, tuberculosis, and paludism	Whole plant extract	[44]
<i>Cyperus esculentus</i> L.	Oaxaca, Santa María Tecomavaca	Roots	Depression	Root extracts	[45]
<i>Cyperus flavescens</i> L.	Oaxaca, Santa María Tecomavaca	Roots	Depression	Root extracts	[45]

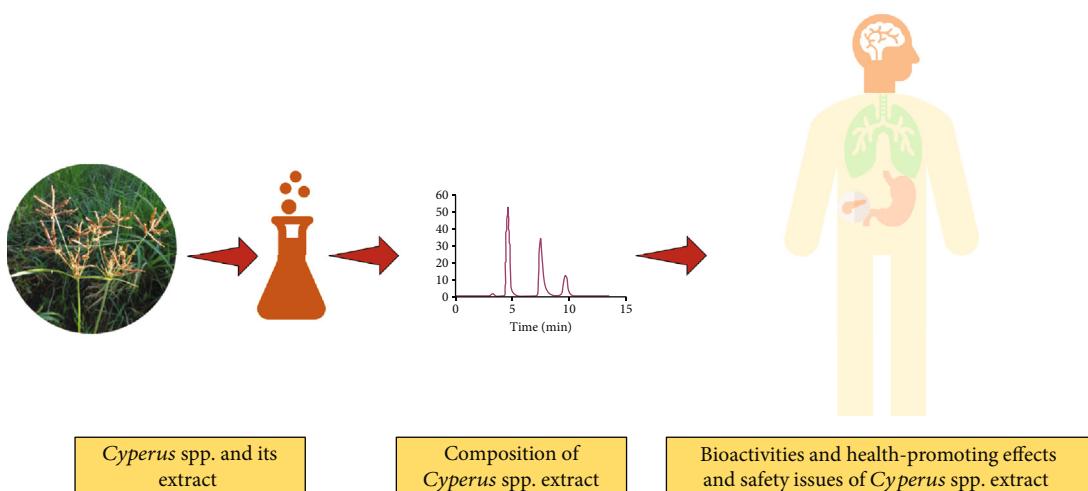


FIGURE 1: Diagram showing various components discussed in the review.

articulatus include α -cyperone, α -corymbolol, α -pinene, caryophyllene oxide, cyperotundone, and mustakone. Researchers also identified articulone, myrtenal, and myrtenol from volatile oil of Nigerian *C. articulatus* [55]. Cameroonian *C. articulatus* hexane extracts displayed the presence of isopatchoul-4(5)en-3-one, mandassidione, mustakone, and almost all sesquiterpene diketones [56]. Similar compounds were also identified in the Brazilian rhizome volatile oil with mustakone (14%), caryophyllene oxide (10.2%), and α -pinene (6.4%) [57]. Volatile oil from *C. articulatus* rhizomes showed the presence of α -pinene (3.5–25.2%), β -

pinene (2.3–12.6%), *trans*-pinocarolol (2.2–5.5%), myrtenal + myrtenol (2.3–5.6%), α -copaene (1.3–2.6%), cyperene (0.7–1.6%), β -selinene (0.8–2.4%), lithol (0.9–5.1%), caryophyllene oxide (3.1–8.3%), mustakone (3.4–9.9%), cyperotundone (2.6–4.1%), and α -cyperone (3.2–8.8%) [58]. These reports suggest a qualitative and quantitative difference in the volatile oil composition. The difference was attributed to various factors like air pollution, altitude, harvesting time, developmental stage, luminosity, seasonality, temperature, water availability, nutrients, UV radiation, and pathogens [59, 60].

TABLE 2: Phytochemicals present in different *Cyperus* species.

Cyperus species	Chemical constituents	Plant part	References
<i>Cyperus articulatus</i> L.	α -Campholenal, α -corymbolol, α -cyperone, α -pinene, cyperol, cyclocolorenone, β -copaen-4- α -ol, p-cymene, caryophyllene oxide, corybolane, cyperotundone, limonene, thuja-2,4(10)-diene, <i>trans</i> -pinocarveol, p-mentha-1,5-dien-8-ol, myrtenal, mustakone	Thick rhizomes	[49, 50]
<i>Cyperus conglomeratus</i> Rottb.	Saponins, steroids, tannins, triterpenes	Whole plant powder	[51]
<i>Cyperus distans</i> L.f.	Artemisia ketone, α -cyperone, cyperene, α -pinene, 1,8-cineole, caryophyllene oxide, endesma-2,4,11-triene, humulene epoxide II, germacrene D, pinocarveol, myrtenol, nor-copernone, zierone	Rhizomes	Lawal et al. [52]
<i>Cyperus esculentus</i> L.	β -Pinene, cymene, cyperene, coumaran, cyperotundone, p-vinylguaiacol, vanillin, cyprotundone	Rhizomes	Gugsa & Yaya [46]
<i>Cyperus longus</i> L.	α -Caryophyllene oxide, β -himachalene, β -caryophyllene oxide, aristolone, humulene oxide, irisone, longiverbenone, viridiflorol	Whole plant powder	Memariani et al. [53]
<i>Cyperus rotundus</i> L.	Isobutyl lactate, thiazol-4(5H)-one-5-(4-nitrobenzylideno)-2-phenyl, <i>cis</i> -pinen-3-ol, <i>trans</i> -p-mentha-2,8-dienol, pyranone, <i>cis</i> -10-nonadecenoic acid, β -santalol, α -copaen-11-ol, β -vatirenene, elema-1,3-dien-6-ol, β -nootkatol, <i>cis</i> -13,16-docasadienoic acid, 25,26-dihydroxy-vitamin D3	—	El-Wakil et al. [54]

2.2. *Cyperus conglomeratus* Rottb. *Cyperus conglomeratus* is a perennial monocot with coarse rhizomes up to 12–16 cm long and 0.2–0.3 cm in width. It is native to India but also grows in temperate, tropical, and subtropical regions [61]. It is a perennial weedy herb commonly found with slim and sheathing leaf base. Phytochemical analysis of several plant extracts revealed the existence of different types of constituents at different amounts with medicinal activities. For example, the crude powder is rich in steroids, while the solvent extract is rich in triterpenes [62]. The therapeutic efficacy was not limited to a specific plant part; every plant part displays a pharmacological activity. Henceforth, the pharmacognostic studies of different plant parts have been performed for different plant organs [63], such as aerial parts, flower, fruit, leaf root, and stem [64–71]. The fatty acid profile of *C. conglomeratus* showed the presence of palmitic, linoleic, heptadecanoic, oleic, myristic, arachidonic, linoeric, and stearic acid. The unsaponifiable fractions of *C. conglomeratus* constituted two other bioactive compounds (β -sitosterol and α -amyrin).

2.3. *Cyperus distans* L.f. *Cyperus distans* is an annual herb of about 0.6–1.5 m tall, mostly found in humid areas along roadsides and rivers and as weeds [72]. The phytochemical study of *C. distans* revealed the presence of scabequinone with antifeeding effects [73]. From 80% of its oil composition, almost 22 compounds were isolated, being zierone (33.8%) the main component. Other noteworthy compounds were caryophyllene oxide (14.1%), α -cyperone (9.1%), humulene epoxide II (3.8%), cyperene (3.2%), endesma-2,4,11-triene (2.9%), nor-copernone (2.9%), and germacrene D (2.8%) [74–77].

2.4. *Cyperus esculentus* L. *Cyperus esculentus*, also known as yellow nutsedge, is a perennial herb growing in tropical and temperate regions of the world. Naturally, it is found as a weed in farming areas and in wastelands [78]. The taste of

tubers is sweet and reported to have health and nutritional benefits [79]. The common names of *C. esculentus* are chufas, earth almond, nutsedge, and rush nut [80]. Cyprotundone, the volatile component p-vinylguaiacol (2-methoxy-4-vinylphenol) [81], and vanillin (4-hydroxy-3-methoxy benzaldehyde) have been identified [82], with interesting bioactive effects that been reported to these biomolecules. As a matter of fact, these molecules are helpful in tracing components in cosmetics, dairy products, drug preparations, and pastry products [83].

2.5. *Cyperus longus* L. *Cyperus longus* is an Egyptian plant, distributed throughout Africa and Europe and to Indian sub-continent and is used as herbal tonic and diuretic [84]. Compounds isolated from this plant are flavonoids, stilbenes, and terpenoids [85–88]. Other compounds from *C. longus* essential oil were also identified such as β -himachalene (46.6%), α -humulene (16.7%), and γ -himachalene (10.1) as main components [86]. In another study, 32 components were identified consisting 83.50% of essential oil using gas chromatography-mass spectroscopy (GC-MS) analysis [53].

2.6. *Cyperus rotundus* L. *Cyperus rotundus* is popularly known as Nagarmotha or purple nutsedge or nut grass [89]. This is a perennial herb with creeping rhizomes 1–3 cm long having a bulbous base. The stems of this herb can attain as the size of about 140 cm, and leaves are grooved on the upper surface. The ethanolic extracts of *C. rotundus* were determined using HPLC, and it was reported to contain two bioactive phenolics, i.e., quercetin and chlorogenic acid [90]. Structures of important members of bioactive compounds from *Cyperus* spp. are shown in Figure 2.

3. Bioactive Effects: Preclinical Evidence

3.1. Antioxidant Activity. Antioxidants are substances which remove reactive species or free radicals from cells and play a

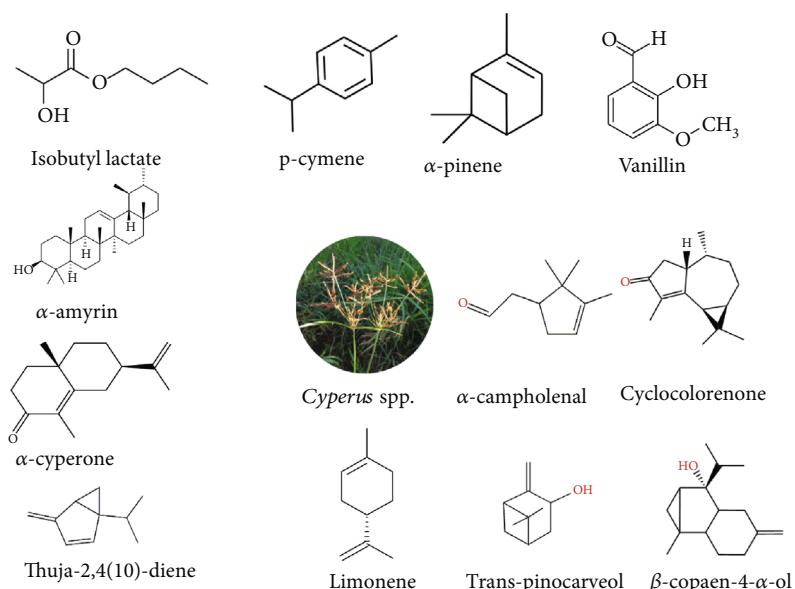


FIGURE 2: Structure of important members of bioactive compounds from *Cyperus* spp.

crucial role in maintaining the health and by preventing the diseases. The antioxidant capacity of *Cyperus* spp. is attributed to the plethora of phytochemicals present. Phenolic compounds, specifically flavonoids, tannins, and coumarins, are present in this species. The presence of these phytochemicals is directly correlated with antioxidant effects [91]. For example, a study assessed the nutritional value, mineral composition, secondary metabolites, and antioxidant activity of 5 wild geophytes: 2 from the Cyperaceae family (*Cyperus capitatus* Vand. and *C. conglomeratus* Rottb.) and 3 from the Poaceae family (*Elymus farctus* (Viv.) Runemark ex Melderis, *Lasiurus scindicus* Henrard, and *Panicum turgidum* Forssk.) collected from the Egyptian coastal desert (Mediterranean coast of the Delta) and the interior desert (Wadi Hagoul). Strong radical scavenging activity with $EC_{50} < 1$ mg/ml assessed using the 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay was reported from the extracts of *C. conglomeratus* and *C. capitatus* [92]. An experiment assessing the antioxidant activity of the extracts of *Cyperus tegetum* Roxb. demonstrated significant DPPH radical, superoxide anion, and hydrogen peroxide scavenging activities compared to the standards, viz., hydroxybutylanisol, butylhydroxytoluene, and ascorbic acid, respectively [93]. In addition, the milk extracted from *C. esculentus* tubers, commonly known as tigernut, was utilized in a study on rats to assess its effect on preventing acetaminophen-induced liver damage (APAP). Its presence led to an increased activity of antioxidant enzyme superoxide dismutase (SOD), while malondialdehyde concentrations were lower than the control group, thus demonstrating a good antioxidant activity [94]. The improvement in the levels of antioxidant enzymes resulted in the decreased reactive species and free radical in acetaminophen-treated liver, thus, managing the situation of liver damage. Similarly, the polyphenolic content of aquatic extracts from *C. rotundus* tubers was found to have protective effects on liver and kidney function caused by

exposure to heavy metals (cadmium chloride) in rats, through scavenging of free radicals [95]. Also, components of the essential oils of *C. articulatus* rhizome encapsulated in chitosan nanoparticles revealed a high potential to eliminate free radicals [96]. The encapsulation improves the stability and also the efficiency of extracts from *Cyperus* spp. resulting in decreasing the stress imposed by free radicals.

Cyperus plant extracts have proven to have a neuroprotective effect caused due to reactive oxygen species (ROS). The deposition of beta-amyloid in the hippocampus promotes oxidative stress, reactive ROS formation, reduction of the antioxidant enzymes activity, and consequently, neuronal death. Previous studies have shown that flavonoids can modulate the function of immune cells, exerting a direct effect against inflammation and oxidative stress [97]. Thus, the antioxidant activity showed by the flavonoids present in *C. rotundus* extracts explains the increase in hippocampal neurogenesis of beta-amyloid in rat models and consequently improves the memory [98]. Orientin, a flavonoid found in *C. esculentus*, decreased oxidative stress generating a neuroprotective effect against cerebral ischemia/reperfusion injury in Sprague-Dawley rats through the middle cerebral artery occlusion method [99].

3.2. Anti-Inflammatory Activity. Numerous studies have proven the potent anti-inflammatory activity of extracts obtained from various plant parts of the *Cyperus* genus [100]. The anti-inflammatory action of the extract from *C. rotundus* rhizome was first described in 1971 [101], and since then, investigations have been done to confirm and understand the anti-inflammatory effect of the different plant parts or active constituents of *C. rotundus*. The compound alpha-cyperone, one of the main phytochemicals found in *C. rotundus* oil, was found to inhibit lipopolysaccharide- (LPS-) stimulated inflammatory response in a murine BV-2 microglial cell line, by activating Akt (protein kinase B)/nuclear

factor-E2-related factor (Nrf)-2/heme oxygenase- (HO-) 1 and suppressing the nuclear factor kappa light chain enhancer of the activated B cell (NF- κ B) pathway [102]. A study concluded that α -cyperone exerts a neuroprotective activity by attenuating the production of inflammatory cytokines in BV-2 cells through activating Akt/Nrf2/HO-1 and suppressing the NF- κ B pathway.

Another study using methanol extracts from *C. rotundus* rhizomes revealed that cyperaline A has high anti-inflammatory activity through inhibition of prostaglandin E2 (PGE-2), cyclooxygenase-2 (COX-2), and arachidonate 5-lipoxygenase (LOX-5) and that sugetriol triacetate, another compound of biological interest in *C. rotundus*, presented a similar effect on PGE-2, COX-2, and LOX-5 enzymes in peripheral blood mononuclear cell (PBMC) lines [103]. α -Cyperone revealed to suppress the inflammatory response in lipopolysaccharide- (LPS-) induced acute lung injury in mice, through inhibiting the growth of inflammatory cells along with cytokines and downregulating the NF- κ B and NLR family pyrin domain containing 3 (NLRP3) signalling pathways [104]. Moreover, recent evidence has shown that the topical application of *C. rotundus* rhizome extract in a rat model with chronic and acute dermatitis leads to a reduction in ear oedema and inflammatory cell infiltration generated by exposure to 12-O-tetradecanoylphorbol-acetate (TPA). This ultimately suggested that the extract could be a potential new therapeutic tool for the treatment of inflammatory skin disorders [90].

3.3. Antimicrobial Activity. The antimicrobial activity of *C. rotundus* extract has been shown in numerous studies [7, 100, 105–109]. In general, it was documented that Gram-positive bacteria were more sensitive to *Cyperus* extracts than Gram-negative bacteria. However, direct comparison of different studies was difficult due to variety of microbiological tests, microbial genera and species, presence of saccharides, herb cultivation conditions, extraction methods, and so on [110]. For example, a study carried out with *C. articulatus* essential oils revealed interesting inhibitory effects on *Staphylococcus aureus* and *Escherichia coli* [96]. Traditional medicine practitioners make use of water primarily as a solvent, but studies have shown that alcohol extracts of plants are much potent and efficacious [111]. The effects of aqueous and alcohol extracts with essential oils from *C. rotundus* tubers on cultures of *Streptococcus mutans*, *Aggregatibacter actinomycetemcomitans*, and *Candida albicans* were investigated. Alcoholic extracts displayed marked inhibition of *S. mutans* and *A. actinomycetemcomitans* growth, making these extracts future candidates for both treatment and prevention of periodontitis and oral cavity affections [105].

Another study used chloroform extracts of *C. conglomeratus*, orally administered to mice, aiming to determine the degree of consumption toxicity. There was no damage to the liver or kidney at the doses used, and a negative impact on the growth of *C. albicans*, *C. dubliniensis*, *C. famata*, *C. glabrata* and *C. inconspicua* was listed. β -Sitosterol and α -amyrin were the most abundant components identified and were the chemicals responsible for the significant effect on the growth of *C. famata* and *C. albicans*, respectively [106].

C. conglomeratus chloroform extracts were also assessed for their antibacterial activity [112]. Extracts demonstrated powerful activity against Gram-positive and Gram-negative bacterial strains, such as *S. aureus*, *Enterococcus aureus*, *Bacillus subtilis*, *E. coli*, and *Pseudomonas aeruginosa*, while butanol and ethyl acetate extracts showed a moderate activity. The antimicrobial activity of *Cyperus* sp. extracts is mainly contributed by the essential oil components. The essential oil components from *Cyperus* spp. easily penetrate inside and create pores in bacterial cells which results in leakage of intracellular components resulting to cell death.

The antimycobacterial activity of *C. rotundus* extracts, evaluated on multidrug-resistant strains of *Mycobacterium tuberculosis*, also revealed to be prominent [113]. In addition, such extracts were used to assess their antibacterial activity and the mode of action against ampicillin-resistant *S. aureus*. The extract revealed a synergistic activity when in combination with ampicillin at the lowest inhibitory concentration. Using electron microscopy, it was observed that the combined treatment induced damages to the peptidoglycans and cell membrane, generating an increase in membrane permeability and revealing an inhibitory activity against β -lactamase [114]. Another study found that the fermented extracts of *C. rotundus* inhibited the growth of *P. aeruginosa*, *B. subtilis*, and *E. coli* [115]. For such reasons, these extracts may be conceived as a natural remedy against infections caused by pathogenic bacteria. More recently, copper oxide nanostructures synthesized using *C. rotundus* extracts revealed an excellent antibacterial activity against *Klebsiella pneumoniae* strains, and the observed inhibitory effects seemed to be associated with factors such as mechanical damage, oxidative damage, and genetic toxicity [116].

3.4. Anticancer Activity. The anticancer activity of *C. rotundus* extracts has also been assessed; the mechanism of action also elucidated the influence on genetic expression. For example, human cervical cancer (HeLa) cell lines exposed to different doses of *C. rotundus* extracts revealed morphological modifications and changes in the degree of chromatin condensation. Microarray analysis also showed that the extract led to the upregulation of 449 genes and downregulation of 484 genes, classified into different interaction pathways, with gene expression induction being associated with apoptosis and cell cycle arrest [117]. The main mechanism of anticancer activity of plant extracts is by inhibiting the cell proliferation or by inducing apoptosis in the cancerous cells. Both of these mechanisms are impaired in the cancerous cells. Plant extracts either halt the cell division or induce apoptosis of cancerous cells by activating the apoptotic factors. *C. rotundus* ethanol extracts were used to evaluate its effects on triple-negative breast cancer cells (TNBC) (negative for estrogen, progesterone receptors, and human epidermal growth factor receptor 2 (HER2) protein overexpression). As main findings, the authors stated that the chemical components present in the extracts inhibited the TNBC cell proliferation, which might be related to cell cycle arrest at the G₀/G₁ phase, thus inducing apoptosis by promoting Bcl-2-associated X protein (Bax) expression and inhibiting B cell lymphoma (Bcl) expression. The n-hexane extract from *C.*

rotundus rhizomes also exhibited an anticancer activity on Michigan Cancer Foundation-7 (MCF-7) breast cancer cell lines, by inducing apoptosis and halting them in G₀-G₁ stages of the cell cycle [118].

The cytotoxic effects of benzoquinones isolated from *Cyperus* sp. roots and tubers were also studied in adenocarcinoma gastric (AGS) and human gastric cancer cell lines. As main achievements, the authors stated that benzoquinones exerted their toxic effect by activating stress in the endoplasmic reticulum, increasing expression of C/EBP homologous protein (CHOP) (mRNA and protein levels), intracellular ROS, changes in calcium dynamics, and caspase-4 activation. The proteasome inhibition caused by hydroxyl cyperaquinone (causing cell death) was first described by the inositol-requiring enzyme 1 α - (IRE1 α -) independent/(PKR-like ER kinase) PERK-dependent pathway in stomach cancer cells [119]. Recently, the anticancer activity was also tested using silver nanoparticles in combination with *C. conglomeratus* extracts. The cytotoxic effect was assessed in MCF-7 breast cancer cells and normal fibroblasts using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT), and a selective cytotoxicity against MCF-7 was stated, while in fibroblasts, no toxic effect was reported. Furthermore, the apoptotic effects were confirmed using annexin V-fluorescein isothiocyanate-propidium iodide (FITC-PI) and real-time PCR for apoptotic genes [120]. Various biological activities of the extracts from *Cyperus* spp. are shown in Figure 3.

3.5. Other Biological Activities. Six sesquiterpenes isolated from *C. rotundus* rhizome methanol extracts were analyzed as an alternative to hormone replacement therapy (HRT). Its estrogenic activity was evaluated in MCF-7 cell lines through its competitive binding to estrogen receptor- (ER- α and ER- β). These isolated compounds revealed to be useful as an alternative to HTR [121]. Tongbi-san (TBS), the name attributed to a set of 3 herbs: *C. rotundus*, *Citrus unshiu*, and *Poria cocos*, has been used in traditional Korean medicine for dysuria. In a study, a significant reduction in body weight and a decrease in weight of epididymal and visceral white adipose tissue were found following oral administration of TBS to male mice (C57BL/6N) for 11 weeks. TBS enhanced the expression of AMP-activated protein kinase (AMPK) and inhibited the expression of transcription factors, such as CCAAT-enhancer-binding proteins (C/EBPs), sterol regulatory element-binding transcription factor 1 (SREBP1), and peroxisome proliferator-activated receptors (PPAR γ) in the liver and white adipose tissue of the epididymis [122].

Cyperus eragrostis seed extracts have also demonstrated vasodilatory properties and ability to inhibit the mammalian arginase enzyme in both an *ex vivo* experiment on rat aortic rings and an *in vitro* assay with purified bovine liver arginase [123]. Through an *in vivo* study in rats, mice, and chicks and *in vitro* study using isolated tissues of the jejunum and ileum of rabbits and rats, the antispasmodic, antidiarrheal, and antiemetic effects of *Cyperus niveus* Retz. were assessed. The presence of flavonoids, phenols, alkaloids, tannins, saponins, and glycosides in the extracts was described to be responsible for the significant inhibition of diarrhea in rats

and for the marked decrease in the intestinal motility of mice [124].

Cyperus articulatus ethanolic extracts revealed to prevent pentylenetetrazol-induced seizures (PTZ) and to increase the gamma aminobutyric acid (GABA) levels in mice with PTZ [125]. Other preclinical (*in vivo* and *in vitro*) were experiments performed with *C. rotundus* to evaluate its effect on gastrointestinal, bronchial, and vascular disorders, as well as pain, emesis, pyrexia, and bacterial infections. A study revealed that the crude extract from *C. rotundus* has remarkable spasmolytic, bronchodilator, and vasodilatory effects, possibly through blockade of calcium channels [126]. A recent study evaluated the antiulcer potential of methanol and ethyl acetate extracts of *Cyperus alternifolius* L. rhizomes and aerial structures in fasted rats with orally administered indomethacin (30 mg/kg). The extracts led to a significant reduction in the number of ulcers and TNF- α content in the stomach. Histopathological examination revealed an improvement in damaged mucosa, with the effect generated by tubers being more effective than that of the control ranitidine [127].

4. Health-Promoting Effects: Data from Clinical Findings

As previously referred (Section 1), *Cyperus* spp. have been used over the years in folk medicine around the world to prevent and even treat different medical afflictions. *C. rotundus* is the most widely used and exploited species from this genus, and the metabolites present in this species are currently well elucidated [128, 129], so as their bioactive effects, analyzed through a number of investigations [130]. For example, the combined effects of *C. rotundus* with other medicinal plants taken as capsules of dehydrated plants or in the form of decoction were found effective against overweight, obesity, Alzheimer's disease (AD), depression, and rheumatoid arthritis. However, no clinical trials on *C. rotundus* used alone could be found.

An Ayurvedic polyherbal formulation, called Trimad, composed of *C. rotundus* tubers, *Embelia ribes* Burm.f. fruits and *Plumbago zeylanica* L. roots traditionally used for the management of overweight and obesity, was investigated by Salunke et al. [131] in 20 overweight and obese individuals. In this clinical study, the authors aimed to assess the effect of the aqueous extract of triphala (formulation made from *Emblica officinalis* L., *Terminalia bellirica* (Gaertn.) Roxb., and *Terminalia chebula* Retz.) (two tablets of 500 mg, twice a day after meals), Trimad, and placebo (dextrin) for the management of obesity and overweight, over 90 days. Significant differences were stated in visceral and subcutaneous fat, as well as other benefits, like positive bowel regulation and a decrease in fatigue in patients who received triphala [131]. Similarly, an Ayurvedic polyherbal mixture (no common name was mentioned) proved to be effective in the management of type 2 diabetes, with findings being compared to metformin effects in a 6-month treatment. The capsules, consisting of *Berberis aristata*, *C. rotundus*, *Cedrus deodara*, *Emblica officinalis*, *Terminalia chebula*, and *Terminalia bellirica*, following HPLC analysis revealed the presence of

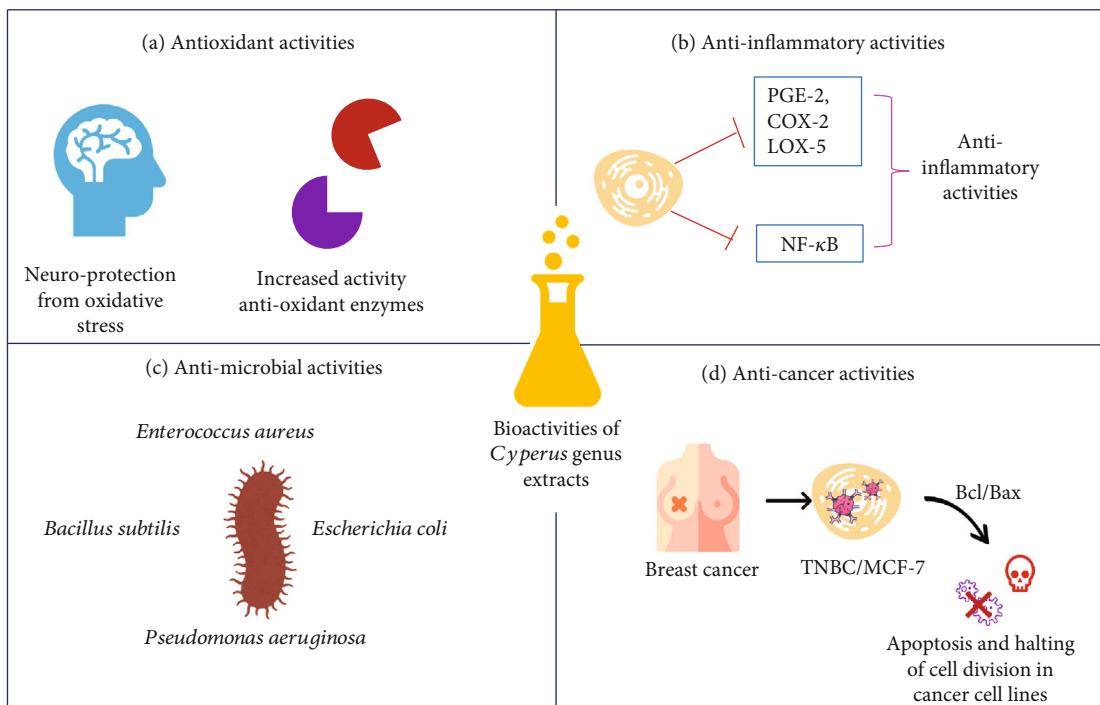


FIGURE 3: Biological activities of the extracts from *Cyperus* spp.

berberine (1.27%), quercetin (0.01%), and gallic acid (3.09%). The randomized study included 93 participants, and 48 of them received 3 g of the herbal treatment for 6 months. The study concluded that the formulation had a positive effect on blood glucose levels, on glycosylated hemoglobin, and on the lipid profile of the patients without showing any adverse effects [132]. Finally, another traditional Ayurvedic formulation with *C. rotundus* [133] was found to be useful for rheumatoid arthritis. The herbomineral formulation (5 g of each plant), consisting of *Vitex negundo*, *C. rotundus*, *Nyctanthes arbor-tristis*, *Smilax glabra*, *Delphinium denudatum*, and *Withania somnifera*, was combined with Maha yogaraj Guggulu Vaiswanar churna and Simhanada Guggulu. No information on its safety or chemical composition was provided in the article, despite the fact that patients (39%) who receive the preparation twice a day before meals for 1 year exhibited a good response to the treatment.

The possible use of *C. rotundus* against neurocognitive disorders has also been reported. The combination of *C. rotundus* with *Crocus sativus* and *Astragalus membranaceus* honey was assessed for its ability to treat a major neurocognitive disorder. A double-blind clinical trial on 60 patients previously diagnosed was performed. The intervention group took two daily capsules of 500 mg each one of the combinations for three months. The results indicated that the combination could be useful to improve the cognitive and depression score. However, the extract used was not characterized, and the preparation method was not indicated [134]. The traditional Iranian herbal medicine, Davaie Loban, composed of *C. rotundus*, *Zingiber officinale*, *Acorus calamus*, *Piper nigrum*, and *Boswellia carterii* has also been tested against AD. In the study, 24 patients with mild to moderate AD took 500 mg of the capsules 3 times daily for 12 weeks.

Results showed that the herbal mixture might have an improvement in the memory of patients. However, despite the safety of the administered mixture which was tested, the preparation method was not detailed [135]. In traditional Chinese medicine, the Guipi decoction, an herbal mixture, is used given its ability to regulate blood pressure and as an antidepressant [136]. So, Zhuang et al. [137] tested the effect of Guipi decoction in 120 elderly patients affected with hypertension and depression. The decoction was prepared as follows: 15 g of a mixture formed by *Codonopsis pilosula*, *Atractylodes*, *Angelica sinensis*, membranous milkvetch root, *Polygon tenuifolia*, and *Arillus longan*; 20 g of *Poria cocos* and *magnesium*; 30 g of calcined *Os Draconis*, calcined oyster shell, light wheat, and *concha haliotidis*; and 10 g of elecampane, fresh ginger, Chinese date, *colla corii asini*, mint, *Albizia julibrissin*, *C. rotundus*, jasmine, and *Coptis chinensis* as well as *spina date seed* of 40 g. The mixture was decocted, and patients received 400 ml of extract for a month. The control group was treated with sertraline. Results concluded that the mixture exerted a curative effect and improved the patients' quality of life by alleviating depression symptoms, although no significant differences were reported in blood pressure.

Some other clinical trials have been performed using the extracts of *Cyperus* spp., but the species investigated were not specified in a large amount of cases. These are the cases of mixtures made with *Cyperus* for sexual dysfunction [138], as anti-inflammatory and analgesic [139] agents and even as lactation inducers [140]. Another critical limitation present in many clinical trials is that in most cases, as previously listed, the herbal mixture was not characterized, and when a chemical characterization was reported, only some polyphenolic compounds were analyzed. In a case, the herbal mixture

was consumed as an aqueous extract, but the polyphenol analysis by HPLC was done in a methanol/water extract. Therefore, the active metabolites responsible for the therapeutic action remain unknown, and the possible synergistic and antagonistic interactions among the plant constituents were not addressed neither described. In addition and also worth noting is that few researches have done a microbiological or safety study on the dehydrated plants when consumed in the form of capsules of dried plants. However, the presence of pathogens, spores, heavy metals, and even aflatoxins was not described in the preparations, which otherwise would have been a potential risk for patients. Moreover, such clinical trials were done with a few patients, and in most of them, both treatment monitoring and fidelity were not precise. Therefore, there is an urgent need to design more robust and reproducible clinical trials to prove the potential health benefits of *Cyperus* spp., which have been widely described via a plethora of *in vitro* and *in vivo* tests.

5. Safety and Adverse Effects

Due to its richness in chemical constituents, *Cyperus* plants have been widely used in folk medicine for multiple afflictions [100, 141, 142]. Thorough screening of literature available on *Cyperus* plants as a popular remedy among various ethnic groups, researchers have increasingly explored their therapeutic potential [143]. However, it is also of extreme interest to evaluate the toxicological aspects of botanical drugs and products for their reliable and safe usage among consumers.

Toxicological data from *C. rotundus* extracts have been reported by several investigators [144]. Most of them reported the use of *Cyperus* extracts as safe [100, 109], with no side effects or even only minor side effects being reported. For example, *C. rotundus* extracts were studied *in vivo* for toxicity and biochemical activities in mice and rats. The lethal dose LD₅₀ of *C. rotundus* root extract, when administered intraperitoneally, was 90 g/kg [145, 146]. Ethanolic extract of dried roots of this plant administered to mice of both sexes showed LD₅₀ > 0.5 mg/kg [147]. Aqua-ethanolic (1:1) extracts of rhizome administered to mice of both sexes produced LD₅₀ of 681.0 mg/kg [148]. In turn, the LD₅₀ of *Cyperus* essential oils was 5000 mg/kg in rats [149]. Other studies reported that a single oral administration of *C. rotundus* ethanolic extract at 5000 mg/kg did not produce signs of toxicity, behavioral changes, mortality, or differences on gross appearance of internal organs in the animals. In subacute toxicity, all rats received a repeated oral dose of 1000 mg/kg of the ethanolic extract for 14 days. The parallel group received the ethanolic extract in the same period but was kept for further 14 days. Application did not produce any effects or reversibility of toxic effects. Thus, it was concluded that the extract did not cause changes in terms of general behaviors, mortality, weight gain, and hematological and clinical blood chemistry parameters in comparison to the control group [150]. Another research also confirmed the safety of *Cyperus* extracts. *C. rotundus* methanolic extracts in mice at doses of 250 and 500 mg/kg body weight showed no toxic effects [151]. On the other side, Lemaire and

coworkers [152] documented that the administration of 45 or 220 mg/kg/day of *C. rotundus* tubers' hexane extracts for 60 days in rats stimulated a significant reduction in weight gain but without toxic effects. Raut and Gaikwad [153] also observed no toxicity symptoms following the administration of *C. rotundus* crude extract at different concentrations and oral doses. Jebasingh et al. [154] performed acute toxicological studies with *C. rotundus* extract and found no mortality or morbidity up to 2000 mg/kg body weight in rats. Toxicity studies also revealed no changes in food, water consumption, and body weight of animals with an increase in white blood cell count and hemoglobin level as well as improvement in kidney and liver function. Krisanapun et al. [155] carried out the acute toxicity test of *C. rotundus* water extracts in rats and reported the single oral LD₅₀ > 5 g/kg body weight. The *C. rotundus* extract used at three doses, 10, 100, and 1000 mg/kg, did not exhibit any sign of toxicity. However, it was observed that at 1000 mg/kg, motor activity was slightly decreased. The effects of *C. rotundus* extract were also assessed on different biochemical parameters (glucose, lipid profile, cardiac enzymes, liver enzymes, and kidney function test). Liver enzymes were found normal, and a nonsignificant increase in serum bilirubin, gamma-glutamyl transferase (GGT), and serum glutamic-pyruvic transaminase (SGPT) was recorded. Also, hematological studies did not show any significant toxic changes, and histopathological examination confirmed that the tested extract was safe and nontoxic [156]. Finally and also worth noting is the use of *Cyperus* spp. for biotechnological purposes, namely, regarding its use as a functional food additive. For example, Carvalho Barros and coworkers [157] evaluated the replacement of beef fat in beef burgers using a tigernut (*C. esculentus*) oil emulsion, to reduce total fat and saturated fatty acids in the studied samples. As main findings, the authors stated that total replacement of beef fat using tigernut oil emulsions in beef burger resulted in a well-accepted and healthier meat product with reduced total and saturated fat contents, as well as increased unsaturated fatty acids [157]. However, further studies are needed to further explore other *Cyperus* sp. agroindustrial and biotechnological potentialities.

6. Conclusions and Perspectives

Across the diverse traditional systems of medicine, *Cyperus* sp. is popularly employed as a potent ethnomedicine owing to its plethora of pharmacological attributes, *viz.*, antioxidant, anti-inflammatory, neuroprotective, antidepressive, antiarthritis, antiobesity, antimicrobial, anticancer, vasodilator, spasmolytic, bronchodilator, and estrogenic properties. This wide array of biological activities is closely linked to the presence of phytochemicals such as α -cyperone, α -corymbolol, α -pinene, caryophyllene oxide, cyperotundone, germacrene D, mustakone, and zierone. However, its wide-ranging use in folk medicine and expansive pharmacological properties are not corroborated with incontrovertible evidences employing animal models, where despite the bioactive phytochemicals of *Cyperus* spp., they have been well-deciphered. Comprehensive investigations on the pharmacological efficacies of isolated compounds are still inadequate.

Moreover, structure-activity analyses on the obtained phytoconstituents have also uncovered the perception of the underlying molecular insights of action of its active extracts and/or phytochemicals. On the other side and although toxicological data have indicated the use of *Cyperus* spp. as safe and effective, conclusive studies encompassing its clinical, toxicological, and safety features are still sparse. In addition and given that reported clinical trials lack herbal mixtures' characterization containing *Cyperus* spp., further studies are needed to explore the precise active constituents. Moreover, the subchronic toxicity as well as their interactions with commonly used conventional drugs to assure a safe and long-term consumption by human subjects also needs further attention of the researchers. Lastly and not least interesting to underline is that, considering the extensive pharmacological potential of *Cyperus* spp., more in-depth research is needed to attain a greater clarity of its mechanism of action.

Conflicts of Interest

The authors declare no conflict of interests.

Authors' Contributions

All authors contributed equally to the manuscript. Conceptualization was performed by J.S.-R., N.C.-M., M.K., W.C.C., and A.D.; validation, investigation, data curation, and writing were performed by all authors; review and editing were performed by J.S.-R., N.C.-M., M.K., W.C.C., and A.D. All the authors read and approved the final manuscript.

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References

- [1] S. R. Sivapalan, "Medicinal uses and pharmacological activities of *Cyperus rotundus* Linn-a review," *International Journal of Scientific and Research Publications*, vol. 3, 2013.
- [2] M. B. Jones, F. Kansiime, and M. J. Saunders, "The potential use of papyrus (*Cyperus papyrus* L.) wetlands as a source of biomass energy for sub-Saharan Africa," *GCB Bioenergy*, vol. 10, no. 1, pp. 4–11, 2018.
- [3] I. Larridon, T. Villaverde, A. R. Zuntini et al., "Tackling rapid radiations with targeted sequencing," *Frontiers in Plant Science*, vol. 1655, 2019.
- [4] J. Dwyer, "*Cyperus Rotundus* L.: An ancient food staple but now designated the world's worst weed," in *Proceedings of 20 th Australasian Weeds Conference*, pp. 251–254, Perth, Western Australia, 2016.
- [5] A. Reznicek, D. Kearns, D. Simpson et al., *Cyperaceae*, Encyclopedia Britannica, 2008.
- [6] L. M. Udari, *Medicinal properties of Cyperus species (sedge family, Cyperaceae)*, 2018.
- [7] A. M. Peerzada, H. H. Ali, M. Naeem, M. Latif, A. H. Bukhari, and A. Tanveer, "*Cyperus rotundus* L.: Traditional uses, phytochemistry, and pharmacological activities," *Journal of Ethnopharmacology*, vol. 174, pp. 540–560, 2015.
- [8] A. M. Peerzada, "Biology, agricultural impact, and management of *Cyperus rotundus* L.: the world's most tenacious weed," *Acta Physiologiae Plantarum*, vol. 39, no. 12, 2017.
- [9] R. Srivastava, A. Singh, and S. Shukla, "Chemical investigation and pharmaceutical action of *Cyperus rotundus*- A review," *Journal of Biologically Active Products from Nature*, vol. 3, no. 3, pp. 166–172, 2013.
- [10] G. Dang, R. Parekar, S. Kamat, A. Scindia, and N. Rege, "Antiinflammatory activity of *Phyllanthus emblica*, *Plumbago zeylanica* and *Cyperus rotundus* in acute models of inflammation," *Phytotherapy Research*, vol. 25, no. 6, pp. 904–908, 2011.
- [11] Y. Coşkuner, R. Ercan, E. Karababa, and A. N. Nazlican, "Physical and chemical properties of chufa (*Cyperus esculentus* L.) tubers grown in the Çukurova region of Turkey," *Journal of the Science of Food and Agriculture*, vol. 82, no. 6, pp. 625–631, 2002.
- [12] E. Sánchez-Zapata, J. Fernández-López, and J. Angel Pérez-Alvarez, "Tiger nut (*Cyperus esculentus*) commercialization: health aspects, composition, properties, and food applications," *Comprehensive Reviews in Food Science and Food Safety*, vol. 11, no. 4, pp. 366–377, 2012.
- [13] U. Shrestha, E. Rosskopf, and D. Butler, "Effect of anaerobic soil disinfection amendment type and C:N ratio on *Cyperus esculentus* sprouting, growth and reproduction," *Weed Research*, vol. 58, no. 5, pp. 379–388, 2018.
- [14] C. N. D. Abbey, *Microbial and sensory properties of canned Tigernut milk*, 2019.
- [15] C. Archer, *Cyperus prolifer Lam.(Cyperaceae)*, 2004.
- [16] A. D. Soren, A. K. Yadav, and E. D. Dhar, "Toxological evaluation of *Cyperus compressus* Linn., a traditionally used anthelmintic plant in India," *Oriental Pharmacy and Experimental Medicine*, vol. 20, no. 3, 2019.
- [17] K. Zaman, "Medicinal plants with hypoglycemic activity," *Journal of Ethnopharmacology*, vol. 26, 1989.
- [18] D. Bora, S. Mehmud, K. K. Das, and H. Medhi, "Report on folklore medicinal plants used for female health care in Assam (India)," *International Journal of Herbal Medicine*, vol. 4, pp. 4–13, 2016.
- [19] M. Ayyanar and S. Ignacimuthu, *Medicinal plants used by the tribals of Tirunelveli hills*, Tamil Nadu to treat poisonous bites and skin diseases, 2005.
- [20] N. Lokendrajit, N. Swapana, C. D. Singh, and C. Singh, "Herbal folk medicines used for urinary and calculi/stone cases complaints in Manipur," *NeBIO*, vol. 2, 2011.
- [21] S. P. Kumar, A. Latheef, and A. Remashree, "Ethnobotanical survey of diuretic and antilithiatic medicinal plants used by the traditional practitioners of Palakkad District," *International Journal of Herbal Medicine*, vol. 2, pp. 52–56, 2014.
- [22] D. Panda, S. S. Rathinayak, and S. Palita, *Crop weeds and its uses in the treatment of common ailments in Koraput district of Odisha, India*, 2015.
- [23] R. Qureshi, G. R. Bhatti, and R. A. Memon, "Ethnomedicinal uses of herbs from northern part of Nara desert, Pakistan," *Pakistan Journal of Botany*, vol. 42, pp. 839–851, 2010.
- [24] L. Dangwal, A. Sharma, N. Kumar, C. Rana, and U. Sharma, "Ethno-medico botany of some aquatic Angiospermae from North-West Himalaya," *Research*, vol. 2, pp. 49–54, 2010.

[25] P. Das and M. Misra, "Some ethnomedicinal plants of Koraput district Orissa," *Ancient Science of Life*, vol. 8, p. 60, 1988.

[26] M. Rao and Y. Varma, "Folklore traditional knowledge on digestive disorders of domestic animals (cattle, sheep and goats) in the Medak district, Telangana, India," *Biolife*, vol. 2, pp. 858–865, 2014.

[27] K. Reddy, G. Trimurthulu, and C. S. Reddy, *Medicinal plants used by ethnic people of Medak district, Andhra Pradesh*, 2010.

[28] F. I. Jahan, M. R. U. Hasan, R. Jahan et al., "A comparison of medicinal plant usage by folk medicinal practitioners of two adjoining villages in Lalmonirhat district, Bangladesh," *American Eurasian Journal of Sustainable Agriculture*, vol. 5, pp. 46–66, 2011.

[29] M. Shahidullah, M. Al-Mujahidee, S. N. Uddin et al., "Medicinal plants of the Santal tribe residing in Rajshahi district, Bangladesh," *American-Eurasian Journal of Sustainable Agriculture*, vol. 3, pp. 220–226, 2009.

[30] J. Suneetha, J. K. Rao, P. P. Rao, and T. S. Reddi, "Ethnomedicine for jaundice by the tribals of East Godavari district, Andhra Pradesh," *Journal of Natural Remedies*, vol. 13, pp. 142–145, 2013.

[31] S. Paul, N. Devi, and G. Sarma, "Ethnobotanical utilization of some medicinal plants by Bodo people of Manas biosphere reserve in the treatment of malaria," *International Research Journal of Pharmacy*, vol. 4, pp. 102–105, 2013.

[32] D. Kumar, Z. A. Bhat, I. A. Chashoo, R. S. Deoda, S. C. Mudgade, and V. Kumar, "Bronchodilator activity in traditional medicines: gift of god kingdom," *Bronchitis*, vol. 171, 2011.

[33] D. K. Holdsworth, "Traditional medicinal plants of Rarotonga, Cook Islands part I," *International Journal of Crude Drug Research*, vol. 28, pp. 209–218, 1990.

[34] P. B. Shamkuwar, A. H. Hoshamani, and I. D. Gonjari, "Antispasmodic effect of *Cyperus rotundus* L (Cyperaceae) in diarrhoea," *Der Pharm Lettre*, vol. 4, p. 522, 2012.

[35] A. Rabelo, I. Oliveira, A. Guimarães et al., "Antinociceptive, anti-inflammatory and antioxidant activities of aqueous extract from *Remirea maritima* (Cyperaceae)," *Journal of Ethnopharmacology*, vol. 145, pp. 11–17, 2013.

[36] A. Moteetee, R. Moffett, and L. Seleteng-Kose, "A review of the ethnobotany of the Basotho of Lesotho and the Free State Province of South Africa (South Sotho)," *South African Journal of Botany*, vol. 122, pp. 21–56, 2019.

[37] S. S. Semenza and A. Maroyi, "Ethnobotanical survey of plants used to treat respiratory infections and related symptoms in the Limpopo province, South Africa," *Journal of Herbal Medicine*, vol. 24, p. 100390, 2020.

[38] A. M. Fakhry and G. S. Aljedaani, "Impact of disturbance on species diversity and composition of *Cyperus conglomeratus* plant community in southern Jeddah, Saudi Arabia," *Journal of King Saud University-Science*, vol. 32, no. 1, pp. 600–605, 2020.

[39] J. Politz and J. Lekeleley, *The knowledge of the Samburu on animal diseases and their traditional methods of treatment*, Unpublished research work, 1988.

[40] J. R. Tabuti, C. B. Kukunda, and P. J. Waako, "Medicinal plants used by traditional medicine practitioners in the treatment of tuberculosis and related ailments in Uganda," *Journal of Ethnopharmacology*, vol. 127, no. 1, pp. 130–136, 2010.

[41] R. Blench and M. Dendo, *Dagomba Plant Names*, Cambridge, United Kingdom, 2006.

[42] H. De Wet and S. Ngubane, "Traditional herbal remedies used by women in a rural community in northern Maputaland (South Africa) for the treatment of gynaecology and obstetric complaints," *South African Journal of Botany*, vol. 94, pp. 129–139, 2014.

[43] S. R. Thornton-Barnett, *Ancestral Pharmacopeias: A Paleoethnobotanical Assessment of Plant Use in the Western Free State, South Africa*, 2013.

[44] M. Razafindraibe, A. R. Kuhlman, H. Rabarison et al., "Medicinal plants used by women from Agnalazaha littoral forest (Southeastern Madagascar)," *Journal of Ethnobiology and Ethnomedicine*, vol. 9, no. 1, p. 73, 2013.

[45] S. L. Guzmán Gutiérrez, R. Reyes Chilpa, and H. Bonilla Jaime, "Medicinal plants for the treatment of "nervios", anxiety, and depression in Mexican Traditional Medicine," *Revista Brasileira de Farmacognosia*, vol. 24, no. 5, pp. 591–608, 2014.

[46] T. Gugsa and E. E. Yaya, "Chemical constituents of the traditional skin care and fragrance nut, *Cyperus esculentus* (tiger-nut)," *American Journal of Essential Oils and Natural Products*, vol. 6, pp. 4–12, 2018.

[47] A. Kumar, A. Niranjan, A. Lehri, R. K. Srivastava, and S. Tewari, "Effect of geographical climatic conditions on yield, chemical composition and carbon isotope composition of nagarmotha (*Cyperus scariosus* R. Br.) essential oil," *Journal of Essential Oil Bearing Plants*, vol. 19, no. 2, pp. 368–373, 2016.

[48] Z. Zhou and H. Zhang, "Phenolic and iridoid glycosides from the rhizomes of *Cyperus rotundus* L," *Medicinal Chemistry Research*, vol. 22, no. 10, pp. 4830–4835, 2013.

[49] M. A. Dikwa, U. A. Abdullahi, S. I. Sadiq et al., "Comparative assessment of antibacterial activities of *Syzygium aromaticum* and *Cyperus articulatus* against *Staphylococcus aureus* and *Escherichia coli*," *Journal of Pharmacy & Bioresources*, vol. 16, no. 2, pp. 139–144, 2019.

[50] N. C. Silva, S. F. Goncalves, L. S. Araújo et al., "In vitro and in vivo antimalarial activity of the volatile oil of *Cyperus articulatus* (Cyperaceae)," *Acta Amazonica*, vol. 49, no. 4, pp. 334–342, 2019.

[51] P. Jyoti, P. Hemali, R. Nilam, and C. Sumitra, *Cyperus conglomeratus (Cyperaceae) a halophyte from Gujarat: physico-chemical, phytochemical and pharmacognostic studies*, 2018.

[52] O. A. Lawal, I. A. Ogunwande, A. R. Opoku, and A. O. Oyedeleji, "Zierone: a sesquiterpene ketone from the essential oil of *Cyperus distans* L. (Cyperaceae)," *Advances in Research*, vol. 6, no. 6, pp. 1–6, 2016.

[53] T. Memariani, T. Hosseini, H. Kamali et al., "Evaluation of the cytotoxic effects of *Cyperus longus* extract, fractions and its essential oil on the PC3 and MCF7 cancer cell lines," *Oncology Letters*, vol. 11, no. 2, pp. 1353–1360, 2016.

[54] E. A. El-Wakil, E. A. Morsi, and H. Abel-Hady, *Phytochemical screening, antimicrobial evaluation and GC-MS analysis of Cyperus rotundus*, 2019.

[55] M. M. Sonwa and W. A. König, "Constituents of the essential oil of *Cyperus alopecuroides*," *Phytochemistry*, vol. 56, no. 4, pp. 321–326, 2001.

[56] B. Nyasse, R. Ghogomu, T. B. Sondengam, M. T. Martin, and B. Bodo, "Mandassidione and other sesquiterpenic ketones from *Cyperus articulatus*," *Phytochemistry*, vol. 27, no. 10, pp. 3319–3321, 1988.

[57] M. D. Zoghbi, E. H. Andrade, J. Oliveira, L. M. Carreira, and G. M. Guilhon, "Yield and chemical composition of the essential oil of the stems and rhizomes of *Cyperus articulatus* L. cultivated in the state of Para, Brazil," *Journal of Essential Oil Research*, vol. 18, pp. 10–12, 2006.

[58] M. D. Zoghbi, E. H. Andrade, L. M. Carreira, and E. A. Rocha, "Comparison of the main components of the essential oils of "priprioca": *Cyperus articulatus* var. *articulatus* L., *C. articulatus* var. *nodosus* L., *C. prolitus* Kunth and *C. rotundus* L," *Journal of Essential Oil Research*, vol. 20, no. 1, pp. 42–45, 2008.

[59] R. P. Adams and A. N. Tashev, "The volatile leaf oils of three *Juniperus communis* varieties from Bulgaria," *Phytologia*, vol. 95, pp. 302–307, 2013.

[60] H. D. Hassanein, N. M. Nazif, A. A. Shahat, F. M. Hamouda, E.-S. A. Aboutable, and M. A. Saleh, "Chemical diversity of essential oils from *Cyperus articulatus*, *Cyperus esculentus* and *Cyperus papyrus*," *Journal of Essential Oil Bearing Plants*, vol. 17, no. 2, pp. 251–264, 2014.

[61] B. B. Ndob, L. E. Mengome, and F. Bivigou, "Ethnobotanical survey of medicinal plants used as anthelmintic remedies in Gabon," *Journal of Ethnopharmacology*, vol. 191, pp. 360–371, 2016.

[62] A. Feizbakhsh and A. Naeemy, "Chemical composition of the essential oil of *Cyperus conglomeratus* Rottb. From Iran," *Journal of Chemistry*, vol. 8, S296 pages, 2011.

[63] G. Tang, X. Lin, X. Lai, X. Gong, and S. Ji, "Pharmacognostic studies of *Psychotria rubra* (Lour.) Poir," *Pharmacog Journal*, vol. 10, 2018.

[64] A. Rabinarayan, J. Switu, C. Rudrappa, and S. Vinay, "Pharmacognostical and phytochemical analysis on leaves of *Homalium ceylanicum* (Gardn.) Benth," *Pharmacognosy Journal*, vol. 10, no. 2, pp. 272–277, 2018.

[65] N. Sharma, S. Singh, and S. K. Singh, "Pharmacognostical standardization and preliminary phytochemical investigations on *Acacia auriculiformis* A. Cunn. Ex. Benth stem bark," *Journal of Medicinal Plants*, vol. 5, pp. 398–402, 2017.

[66] N. K. Deb, A. Singh, D. S. Rathore, G. K. Dash, and J. Deb, "Pharmacognostic studies of the stem bark of *Chloroxylon swietenia* DC," *Indian Journal of Pharmaceutical and Biological Research*, vol. 3, no. 4, pp. 1–5, 2015.

[67] Y. Baravalia, K. Nagani, and S. Chanda, "Evaluation of pharmacognostic and physicochemical parameters of *Woodfordia fruticosa* Kurz. Flowers," *Pharmacognosy Journal*, vol. 2, no. 18, pp. 13–18, 2011.

[68] B. Meghashree, T. Shantha, G. Venkateshwarlu, and S. Bhat, "Comparative pharmacognostical and histochemical studies on *Benincasa Hispida* (Thunb.) Cogn.—fruit and seed," *International Journal of Herbal Medicine*, vol. 5, pp. 17–24, 2017.

[69] J. Pande, H. Padalia, S. Donga, and S. Chanda, "Development of quality control parameters for the standardization of *Aegle marmelos* (Roxb) seed," *International Journal of Pharmaceutical Sciences and Research*, vol. 9, pp. 2387–2394, 2018.

[70] R. P. Yejella, "Pharmacognostic study of *Argyreia pilosa* Wight & Arn. root," *Journal of Pharmaceutical & Health Sciences*, vol. 5, pp. 207–216, 2017.

[71] A. Yazdinezhad, N. Ramezanloo, and S. Mozaffari, *Pharmacognostic and phytochemical investigation of *Heracleum persicum* Desf. ex Fischer*, 2016.

[72] P. Goetghebeur, "Cyperaceae," in *Flowering Plants: Monocotyledons*, pp. 141–190, Springer, 1998.

[73] M. Morimoto, Y. Fujii, and K. Komai, "Antifeedants in Cyperaceae: coumaran and quinones from *Cyperus* spp.," *Phytochemistry*, vol. 51, no. 5, pp. 605–608, 1999.

[74] O. A. Lawal and A. O. Oyedele, "The composition of the essential oil from *Cyperus distans* Rhizome," *Natural product communications*, vol. 4, no. 8, p. 1934578X0900400, 2009.

[75] O. A. Lawal and A. O. Oyedele, "Chemical composition of the essential oils of *Cyperus rotundus* L. from South Africa," *Molecules*, vol. 14, no. 8, pp. 2909–2917, 2009.

[76] O. A. Lawal, I. A. Ogunwande, A. R. Opoku, and A. O. Oyedele, "Chemical composition and antibacterial activity of essential oils from the rhizomes of *Cyperus papurus* L., grown in South Africa," *Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas*, vol. 15, pp. 136–143, 2016.

[77] O. A. Lawal, A. B. Ojekale, O. S. Oladimeji et al., "Antioxidant activity, total phenolic and flavonoid contents of essential oils of three *Cyperus* species (Cyperaceae)," *Journal of Pharmaceutical Research International*, vol. 7, pp. 52–62, 2015.

[78] T. Sunday, "Isolation and physicochemical characterization of tigernut (*Cyperus esculentus*) starch as a potential industrial biomaterial," *International Journal of materials science and applications*, vol. 3, p. 37, 2014.

[79] M. Venkatachalam and S. K. Sathe, "Chemical composition of selected edible nut seeds," *Journal of Agricultural and Food Chemistry*, vol. 54, no. 13, pp. 4705–4714, 2006.

[80] J. Rubert, A. Monforte, K. Hurkova et al., "Untargeted metabolomics of fresh and heat treatment Tiger nut (*Cyperus esculentus* L.) milks reveals further insight into food quality and nutrition," *Journal of Chromatography A*, vol. 1514, pp. 80–87, 2017.

[81] H. J. Hussein, M. Y. Hadi, and I. H. Hameed, "Study of chemical composition of *Foeniculum vulgare* using Fourier transform infrared spectrophotometer and gas chromatography - mass spectrometry," *Journal of Pharmacognosy and Phytotherapy*, vol. 8, no. 3, pp. 60–89, 2016.

[82] V. T. Karathanos, I. Mountzinos, K. Yannakopoulou, and N. K. Andrikopoulos, "Study of the solubility, antioxidant activity and structure of inclusion complex of vanillin with β -cyclodextrin," *Food Chemistry*, vol. 101, no. 2, pp. 652–658, 2007.

[83] W. Wong, Y. Zhou, X. Huang, and Q. Zhang, "Preparative isolation and purification of cyperotundone and α -cyperone from *Cyperus rotundus* Linn. with high speed counter-current chromatography," *International Journal of Medicinal and Aromatic Plants*, vol. 3, pp. 163–168, 2013.

[84] T. Morikawa, F. Xu, H. Matsuda, and M. Yoshikawa, "Structures of novel norstilbene dimer, longusone A, and three new stilbene dimers, longusols A, B, and C, with antiallergic and radical scavenging activities from Egyptian natural medicine *Cyperus longus*," *Chemical and Pharmaceutical Bulletin*, vol. 58, no. 10, pp. 1379–1385, 2010.

[85] J. Harborne, "Distribution and taxonomic significance of flavonoids in the leaves of the Cyperaceae," *Phytochemistry*, vol. 10, no. 7, pp. 1569–1574, 1971.

[86] A. Ait-Ouazzou, S. Lorán, A. Arakrak et al., "Evaluation of the chemical composition and antimicrobial activity of *Mentha pulegium*_, *Juniperus phoenicea*_, and *Cyperus longus*_ essential oils from Morocco," *Food Research International*, vol. 45, no. 1, pp. 313–319, 2012.

[87] F. Xu, T. Morikawa, H. Matsuda, K. Ninomiya, and M. Yoshikawa, "Structures of new sesquiterpenes and hepatoprotective constituents from the Egyptian herbal medicine *Cyperus longus*," *Journal of Natural Products*, vol. 67, no. 4, pp. 569–576, 2004.

[88] R. Sharma and R. Gupta, "Cyperus rotundus extract inhibits acetylcholinesterase activity from animal and plants as well as inhibits germination and seedling growth in wheat and tomato," *Life Sciences*, vol. 80, no. 24–25, pp. 2389–2392, 2007.

[89] M. M. Sonwa and W. A. König, "Chemical study of the essential oil of *Cyperus rotundus*," *Phytochemistry*, vol. 58, no. 5, pp. 799–810, 2001.

[90] F. G. Rocha, M. Brandenburgde, P. L. Pawloski et al., "Pre-clinical study of the topical anti-inflammatory activity of *Cyperus rotundus* L. extract (Cyperaceae) in models of skin inflammation," *Journal of Ethnopharmacology*, vol. 254, article 112709, 2020.

[91] S. Kilani-Jaziri, W. Bhouri, I. Skandiani, I. Limem, L. Chekir-Ghedira, and K. Ghedira, "Phytochemical, antimicrobial, antioxidant and antigenotoxic potentials of *Cyperus rotundus* extracts," *South African Journal of Botany*, vol. 77, no. 3, pp. 767–776, 2011.

[92] S. L. Al-Rowaily, A. M. Abd-Elgawad, S. M. Alghanem, W. A. A. Al-Taisan, and Y. A. El-Amier, "Nutritional value, mineral composition, secondary metabolites, and antioxidant activity of some wild geophyte sedges and grasses," *Plants*, vol. 8, no. 12, p. 569, 2019.

[93] A. Chatterjee, R. Khanra, and P. M. A. J. I. Chakraborty, "Phytochemical investigation and evaluation of *in vitro* antioxidant activity of the plant *Cyperus tegetum* Roxb," *Asian Journal of Pharmaceutical and Clinical Research*, vol. 12, no. 11, pp. 18–23, 2019.

[94] N. O. Onuoha, N. O. Ogbusa, A. N. Okorie, and C. E. Ejike, "Tigernut (*Cyperus esculentus* L.) milk' as a potent nutri-drink' for the prevention of acetaminophen-induced hepatotoxicity in a murine model," *Journal of Intercultural Ethnopharmacology*, vol. 6, no. 3, p. 290, 2017.

[95] A. M. A. Aldulaimi and F. F. Husain, "Effect of aqueous extract *Cyperus rotundus* tubers as antioxidant on liver and kidney functions in albino males rats exposed to cadmium chloride toxic," *Baghdad Science Journal*, vol. 16, no. 2, article 0315, 2019.

[96] D. Kavaz, M. Idris, and C. Onyebuchi, "Physiochemical characterization, antioxidative, anticancer cells proliferation and food pathogens antibacterial activity of chitosan nanoparticles loaded with *Cyperus articulatus* rhizome essential oils," *International Journal of Biological Macromolecules*, vol. 123, pp. 837–845, 2019.

[97] R. S. Dhillon, S. Singh, S. Kundra, and A. S. Basra, "Studies on the chemical composition and biological activity of essential oil from *Cyperus rotundus* Linn," *Plant Growth Regulation*, vol. 13, no. 1, pp. 89–93, 1993.

[98] Z. Shakerin, E. Esfandiari, M. Ghanadian, S. Razavi, H. Alaei, and G. Dashti, "Therapeutic effects of *Cyperus rotundus* rhizome extract on memory impairment, neurogenesis and mitochondria in beta-amyloid rat model of Alzheimer's disease," *Metabolic Brain Disease*, vol. 35, no. 3, pp. 451–461, 2020.

[99] S. Q. Jing, S. S. Wang, R. M. Zhong et al., "Neuroprotection of *Cyperus esculentus* L. orientin against cerebral ischemia/reperfusion induced brain injury," *Neural Regeneration Research*, vol. 15, no. 3, pp. 548–556, 2020.

[100] A. E. Al-Snafi, "A review on *Cyperus rotundus* A potential medicinal plant," *IOSR Journal of Pharmacy*, vol. 6, pp. 32–48, 2016.

[101] M. B. Gupta, T. K. Palit, N. Singh, and K. P. Bhargava, "Pharmacological studies to isolate the active constituents from *Cyperus rotundus* possessing anti-inflammatory, anti-pyretic and analgesic activities," *The Indian Journal of Medical Research*, vol. 59, no. 1, pp. 76–82, 1971.

[102] B. Huang, D. He, G. Chen et al., "α-Cyperone inhibits LPS-induced inflammation in BV-2 cells through activation of Akt/Nrf2/HO-1 and suppression of the NF-κB pathway," *Food and Function*, vol. 9, no. 5, pp. 2735–2743, 2018.

[103] S. R. Mohamed, G. A. A. Khayat, M. T. Zayed, and M. F. S. El-Kholy, "Anti-inflammatory terpenoids from *Cyperus rotundus* rhizomes," *Pakistan Journal of Pharmaceutical Sciences*, vol. 31, no. 4, pp. 1449–1456, 2018.

[104] X. Liu, X. Jin, D. Yu, and G. Liu, "Suppression of NLRP3 and NF-κB signaling pathways by α-Cyperone via activating SIRT1 contributes to attenuation of LPS-induced acute lung injury in mice," *International Immunopharmacology*, vol. 76, no. August, p. 105886, 2019.

[105] M. Khojaste, M. Yazdanian, E. Tahmasebi, M. Shokri, B. Houshmand, and R. Shahbazi, "Cell toxicity and inhibitory effects of *Cyperus rotundus* extract on *Streptococcus mutans*, *Aggregatibacter actinomycetemcomitans* and *Candida albicans*," *European Journal of Translational Myology*, vol. 28, no. 4b, p. 7917, 2018.

[106] G. H. Al-Hazmi, A. S. Awaad, M. R. Alothman, and S. I. Alqasumi, "Anticandidal activity of the extract and compounds isolated from *Cyperus conglomeratus* Rottb," *Saudi Pharmaceutical Journal*, vol. 26, no. 6, pp. 891–895, 2018.

[107] S. K. Sharma and A. P. Singh, "Morphological, microscopical and physico-chemical investigations on the rhizomes of *Cyperus rotundus* Linn," *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, vol. 2, no. 3, pp. 798–806, 2011.

[108] M. P. Prasad, "Analysis of antimicrobial compounds in *Cyperus rotundus* and *Azadirachta indica* against human pathogens," *International Journal of Current Microbiology and Applied Sciences*, vol. 3, no. 3, pp. 206–210, 2014.

[109] A. S. Kabbashi, S. E. A. Mohammed, A. Z. Almagboul, and I. F. Ahmed, "Antimicrobial activity and cytotoxicity of ethanolic extract of *Cyperus rotundus* L," *American Journal of Pharmacy and Pharmaceutical Sciences*, vol. 2, pp. 1–13, 2015.

[110] R. Haghgoo, M. Mehran, H. F. Zadeh, E. Afshari, and N. F. Zadeh, "Comparison between antibacterial effect of chlorhexidine 0.2% and different concentrations of *Cyperus rotundus* extract: An *in vitro* study," *Journal of International Society of Preventive and Community Dentistry*, vol. 7, no. 5, pp. 242–246, 2017.

[111] H. J. de Boer, A. Kool, A. Broberg, W. R. Mziray, I. Hedberg, and J. J. Levenfors, "Anti-fungal and anti-bacterial activity of some herbal remedies from Tanzania," *Journal of Ethnopharmacology*, vol. 96, no. 3, pp. 461–469, 2005.

[112] B. Guessoum, A. Hadj Seyd, A. Kemassi, and O. Rahim, "Évaluation de l'activité antibactérienne du *Cyperus conglomeratus* (Cyperaceae)," *Phytothérapie*, vol. 19, 2021.

[113] V. K. Gupta, A. Kaushik, D. S. Chauhan, R. K. Ahirwar, S. Sharma, and D. Bisht, "Anti-mycobacterial activity of some medicinal plants used traditionally by tribes from Madhya Pradesh, India for treating tuberculosis related symptoms," *Journal of Ethnopharmacology*, vol. 227, pp. 113–120, 2018.

[114] P. Cheypratub, W. Leeansaksiri, and G. Eumkeb, "The Synergy and Mode of Action of *Cyperus rotundus* L. Extract Plus Ampicillin against Ampicillin-Resistant *Staphylococcus aureus*," *Evidence-Based Complementary and Alternative Medicine*, vol. 2018, Article ID 3438453, 11 pages, 2018.

[115] H. Kadum, "Metabolomic profiling elucidated by 1h- NMR and the correlation with antibacterial and antioxidant activity of (*Cyperus rotundus* L) fermented by lactic acid bacteria," *Journal of Pure and Applied Microbiology*, vol. 13, no. 3, pp. 1475–1482, 2019.

[116] S. Suresh, R. Ilakiya, G. Kalaiyan et al., "Green Synthesis of Copper Oxide Nanostructures using Cynodon dactylon and *Cyperus rotundus* Grass Extracts for Antibacterial Applications," *Ceramics International*, vol. 46, no. 8, pp. 12525–12537, 2020.

[117] C. H. Lin, S. F. Peng, F. S. Chueh, Z. Y. Cheng, C. L. Kuo, and J. G. Chung, "The Ethanol Crude Extraction of *Cyperus Rotundus* Regulates Apoptosis-associated Gene Expression in HeLa Human Cervical Carcinoma Cells In Vitro," *Anticancer Research*, vol. 39, no. 7, pp. 3697–3709, 2019.

[118] D. Simorangkir, M. Masfria, U. Harahap, and D. Satria, "Activity anticancer n-hexane fraction of *Cyperus rotundus* L. Rhizome to breast cancer MCF-7 cell line," *Open Access Macedonian Journal of Medical Sciences*, vol. 7, no. 22, pp. 3904–3906, 2019.

[119] V. Ribeiro, P. B. Andrade, P. Valentão, and D. M. Pereira, "Benzoquinones from *Cyperus* spp. trigger IRE1 α -independent and PERK- dependent ER stress in human stomach cancer cells and are novel proteasome inhibitors," *Phytomedicine*, vol. 63, article 153017, 2019.

[120] A. G. Al-Nuairi, K. A. Mosa, M. G. Mohammad, A. El-Keblawy, S. Soliman, and H. Alawadhi, "Biosynthesis, Characterization, and Evaluation of the Cytotoxic Effects of Biologically Synthesized Silver Nanoparticles from *Cyperus conglomeratus* Root Extracts on Breast Cancer Cell Line MCF-7," *Biological Trace Element Research*, vol. 194, no. 2, pp. 560–569, 2020.

[121] Y. J. Park, H. Zheng, J. H. Kwak, and K. H. Chung, "Sesquiterpenes from *Cyperus rotundus* and 4 α ,5 α -oxidoeudesm-11-en-3-one as a potential selective estrogen receptor modulator," *Biomedicine and Pharmacotherapy*, vol. 109, pp. 1313–1318, 2019.

[122] Y. J. Park, G. S. Lee, S. Y. Cheon, Y. Y. Cha, and H. J. An, "The anti-obesity effects of Tongbi-san in a high-fat diet-induced obese mouse model," *BMC Complementary and Alternative Medicine*, vol. 19, no. 1, pp. 1–14, 2019.

[123] K. Arraki, P. Totoson, A. Decendit et al., "Cyperaceae Species Are Potential Sources of Natural Mammalian Arginase Inhibitors with Positive Effects on Vascular Function," *Journal of Natural Products*, vol. 80, no. 9, pp. 2432–2438, 2017.

[124] A. Aleem and K. H. Janbaz, "Dual mechanisms of anti-muscarinic and Ca++ antagonistic activities to validate the folkloric uses of *Cyperus niveus* Retz. as antispasmodic and antidiarrheal," *Journal of Ethnopharmacology*, vol. 213, pp. 138–148, 2018.

[125] L. K. Herrera-Calderon, O. Santiváñez-Acosta, R. Pari-Olarte, B. Enciso-Roca, E. C. Montes, and V. M. L. A. Acevedo, "Anticonvulsant effect of ethanolic extract of *Cyperus articulatus* L. leaves on pentylenetetrazol induced seizure in mice," *Journal of Traditional and Complementary Medicine*, vol. 8, no. 1, pp. 95–99, 2018.

[126] M. Hussain, H. M. Waqas, I. Hussain, A. Majeed, S. M. Raza, and K. H. Janbaz, "Pharmacological validation of the folkloric uses of *Cyperus rotundus* L. In different ailments: An *in vivo* and *in vitro* research," *Pakistan Journal of Pharmaceutical Sciences*, vol. 31, no. 1, pp. 95–102, 2018.

[127] A. R. H. Farrag, H. M. I. Abdallah, A. R. Khattab et al., "Anti-ulcer activity of *Cyperus alternifolius* in relation to its UPLC-MS metabolite fingerprint: A mechanistic study," *Phytomedicine*, vol. 62, article 152970, 2019.

[128] F. I. Nuryana, M. A. Chozin, and D. Guntoro, "High-Performance Liquid Chromatography analysis for α -cyperone and nootkatone from the tuber of nutsedge (*Cyperus rotundus* L.) in the tropics," *Rasayan Journal of Chemistry*, vol. 12, no. 1, pp. 360–365, 2019.

[129] F. Li, Y. Zhang, X. Wei, C. Song, M. Qiao, and H. Zhang, "Metabolic profiling of Shu-Yu capsule in rat serum based on metabolic fingerprinting analysis using HPLC-ESI-MSn," *Molecular Medicine Reports*, vol. 13, pp. 4191–4204, 2016.

[130] A. Kamala, S. K. Middha, and C. S. Karigar, "Plants in traditional medicine with special reference to *Cyperusrotundus* L.: a review," *Biotechnology*, vol. 8, no. 7, p. 309, 2018.

[131] M. Salunke, J. Banjare, and S. Bhalerao, "Effect of selected herbal formulations on anthropometry and body composition in overweight and obese individuals: A randomized, double blind, placebo-controlled study," *Journal of Herbal Medicine*, vol. 17–18, p. 100298, 2019.

[132] H. Awasthi, R. Nath, K. Usman et al., "Effects of a standardized *Ayurvedic* formulation on diabetes control in newly diagnosed Type-2 diabetics; a randomized active controlled clinical study," *Complementary Therapies in Medicine*, vol. 23, no. 4, pp. 555–561, 2015.

[133] A. Rakesh and P. L. Soni, "Clinical evaluation of an Ayurvedic formulation in management of rheumatoid arthritis," *Biochemical and Cellular Archives*, vol. 10, no. 2, pp. 185–189, 2010.

[134] S. Akouchekian, V. Omranifard, M. R. Maracy, A. Pedram, and A. A. Zefreh, "Efficacy of herbal combination of sedge, saffron, and *Astragalus* honey on major neurocognitive disorder," *Journal of Research in Medical Sciences: The Official Journal of Isfahan University of Medical Sciences*, vol. 23, no. 1, p. 58, 2018.

[135] H. Tajadini, R. Saifadini, R. Choopani, M. Mehrabani, M. Kamalinejad, and A. A. Haghdoost, "Herbal medicine Davaie Loban in mild to moderate Alzheimer's disease: A 12-week randomized double-blind placebo-controlled clinical trial," *Complementary Therapies in Medicine*, vol. 23, no. 6, pp. 767–772, 2015.

[136] C. Sheng, Z. Chen, H. Cui et al., "Is the Chinese medicinal formula Guipi Decoction effective as an adjunctive treatment for depression? A meta-analysis of randomized controlled trials," *Chinese Journal of Integrative Medicine*, vol. 23, no. 5, pp. 386–395, 2017.

[137] X. Zhuang, J. Yang, and L. Sun, "Effect of add-substract of Guipi decoction on blood pressure patient and the quality of life upon treatment on senile hypertension with depression," *Acta Medica Mediterranea*, vol. 34, pp. 273–277, 2018.

[138] L. Lai, A. Flower, M. Moore, P. Prescott, and G. Lewith, "Polycystic Ovary syndrome: A Randomised feasibility and pilot study using Chinese Herbal medicine to explore Impact on Dysfunction (ORCHID)—Study protocol," *European Journal of Integrative Medicine*, vol. 6, no. 3, pp. 392–399, 2014.

[139] M. I. Thabrew, M. G. Dharmasiri, and L. Senaratne, "Anti-inflammatory and analgesic activity in the polyherbal formulation Maharasnadhi Quathar," *Journal of Ethnopharmacology*, vol. 85, no. 2, pp. 261–267, 2003.

[140] T. Ushiroyama, K. Sakuma, H. Souen et al., "Xiong-gui-tiao-xue-yin (Kyuki-chouketsu-in), a Traditional Herbal Medicine, Stimulates Lactation with Increase in Secretion of Prolactin but not Oxytocin in the Postpartum Period," *The American Journal of Chinese Medicine*, vol. 35, no. 2, pp. 195–202, 2007.

[141] K. J. Soumaya, M. Dhekra, C. Fadwa et al., "Pharmacological, antioxidant, genotoxic studies and modulation of rat splenocyte functions by *Cyperus rotundus* extracts," *BMC Complementary and Alternative Medicine*, vol. 13, no. 1, p. 28, 2013.

[142] K. J. Soumaya, M. Dhekra, C. Fadwa et al., "Pharmacological, antioxidant, genotoxic studies and modulation of rats plenocyte functions by *Cyperus rotundus* extracts," *BMC Complementary and Alternative Medicine*, vol. 13, p. 28, 2014.

[143] A. K. Meena, A. K. Yadav, U. S. Niranjan, B. Singh, A. K. Nagariya, and M. Verma, "Review on *Cyperus rotundus* - A potential herb," *International Journal of Pharmaceutical and Clinical Research*, vol. 2, no. 1, pp. 20–22, 2010.

[144] P. Dhar, D. G. Dhar, A. K. S. Rawat, and S. Srivastav, "Medicinal chemistry and biological potential of *Cyperus rotundus* Linn.: An overview to discover elite chemotype(s) for industrial use," *Industrial Crops and Products*, vol. 108, pp. 232–247, 2017.

[145] B. A. Akperbekova and R. A. Abdullaev, "Diuretic effect of drug form and galenicals from the roots of *Cyperus rotundus* growing in Azerbaijan," *Izv Akad Nauk AzSSR Ser Biol Nauk*, vol. 4, p. 98, 1966.

[146] B. A. Akperbekova and D. Y. Guscinov, "Studies on the influence of pharmaceutical preparations from rhizomes of *Cyperus rotundus* growing in Azerbaijan on the heart and vascular system," *Azerbaidzhanskii Meditsinskii Zhurnal*, vol. 43, no. 7, pp. 12–17, 1966.

[147] W. S. Woo, E. B. Lee, and I. Chang, "Biological evaluation of Korean medicinal plants II," *Yakhak Hoe Chi*, vol. 21, pp. 177–183, 1977.

[148] B. N. Dhawan, M. P. Dubey, B. N. Mehrotra, R. P. Rastogi, and J. S. Tandon, "Screening of Indian plants for biological activity. Part 9," *Indian Journal of Experimental Biology*, vol. 18, pp. 594–606, 1980.

[149] S. Biradar, V. A. Kangarkar, Y. Mandavkar, M. Thakur, and N. Chougule, "Anti-inflammatory, anti-arthritic, analgesic and anticonvulsant activity of *Cyperus* essential oils," *International Journal of Pharmacy and Pharmaceutical Sciences*, vol. 2, pp. 112–115, 2010.

[150] S. Thanabhorn, K. Jaijoy, S. Thamaree, K. Ingkaninan, and A. Panthong, "Acute and subacute toxicities of the ethanol extract from the rhizomes of *Cyperus rotundus* Linn," *Mahidol University Journal of Pharmaceutical Sciences*, vol. 32, no. 1-2, pp. 15–22, 2005.

[151] S. J. Uddin, K. Mondal, J. A. Shilpi, and M. T. Rahman, "Anti-diarrhoeal activity of *Cyperus rotundus*," *Fitoterapia*, vol. 77, pp. 134–136, 2006.

[152] B. Lemaire, A. Touché, I. Zbinden et al., "Administration of *Cyperus rotundus* tubers extract prevents weight gain in obese Zuckerrats," *Phytotherapy Research*, vol. 21, pp. 724–730, 2007.

[153] N. A. Raut and N. J. Gaikwad, "Antidiabetic potential of fractions of hydro-ethanol extract of *Cyperus rotundus* L. (Cyperaceae). Research Journal of Pharmaceutical," *Biological and Chemical Sciences*, vol. 3, pp. 1014–1019, 2012.

[154] D. Jebasingh, D. D. Jackson, S. Venkataraman, and B. S. Emerald, "Physiochemical and toxicological studies of the medicinal plant *Cyperus rotundus* L (Cyperaceae)," *International Journal of Applied Research in Natural Products*, vol. 5, pp. 1–8, 2012.

[155] C. Krisanapun, Y. Wongkrajang, R. Temsiririrkkul, B. Kongsaktragoon, and P. Peungvicha, "Anti-diabetic effect and acute toxicity of the water extract of *Cyperus rotundus* L. in rats," *The FASEB Journal*, vol. 26, no. S1, pp. 686–688, 2012.

[156] M. Ahmad, M. Mahayrookh, A. B. Rehman, and N. Jahan, "Analgesic, antimicrobial and cytotoxic effect of *Cyperus rotundus* ethanolic extract," *Pakistan Journal of Pharmacology*, vol. 29, no. 2, pp. 7–13, 2012.

[157] J. Carvalho Barros, P. E. S. Munekata, F. A. L. de Carvalho et al., "Use of tiger nut (*Cyperus esculentus* L.) oil emulsion as animal fat replacement in beef burgers," *Foods*, vol. 9, no. 1, p. 44, 2020.